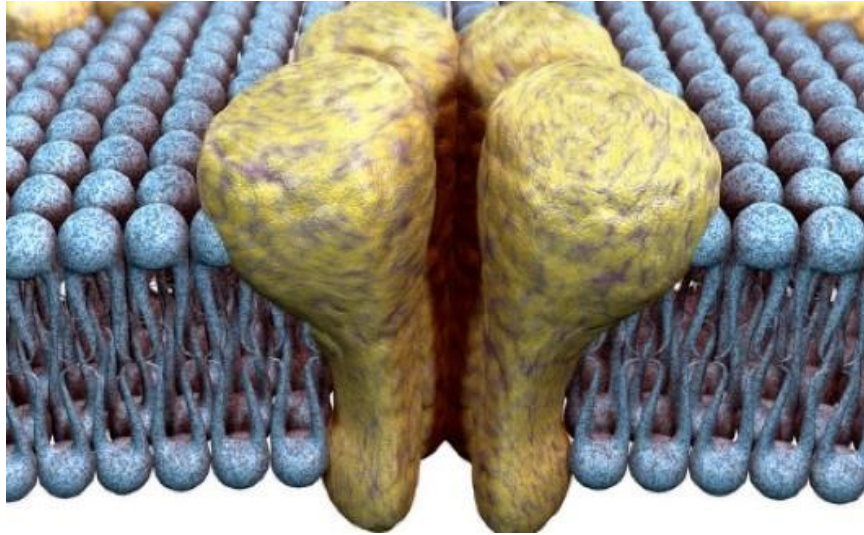




Up/down and left/right by the heart transcriptome

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NELEN Rm.369, tel 936-261-9926, daiacobas@pvamu.edu

Channelopathies: molecular and genetic mechanisms



Chapter 1: Drug discovery, tools and theory

www.nature.com/scientificreports

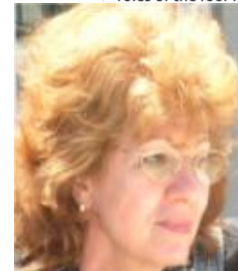
scientific reports

Check for updates

OPEN Transcriptomic uniqueness and commonality of the ion channels and transporters in the four heart chambers

Sanda Iacobas¹, Bogdan Amuzescu² & Dumitru A. Iacobas^{3,4}✉

Myocardium transcriptomes of left and right atria and ventricles from four adult male C57Bl/6j mice were profiled with Agilent microarrays to identify the differences responsible for the distinct functional roles of the four heart chambers. Female mice were not investigated owing to their transcriptome

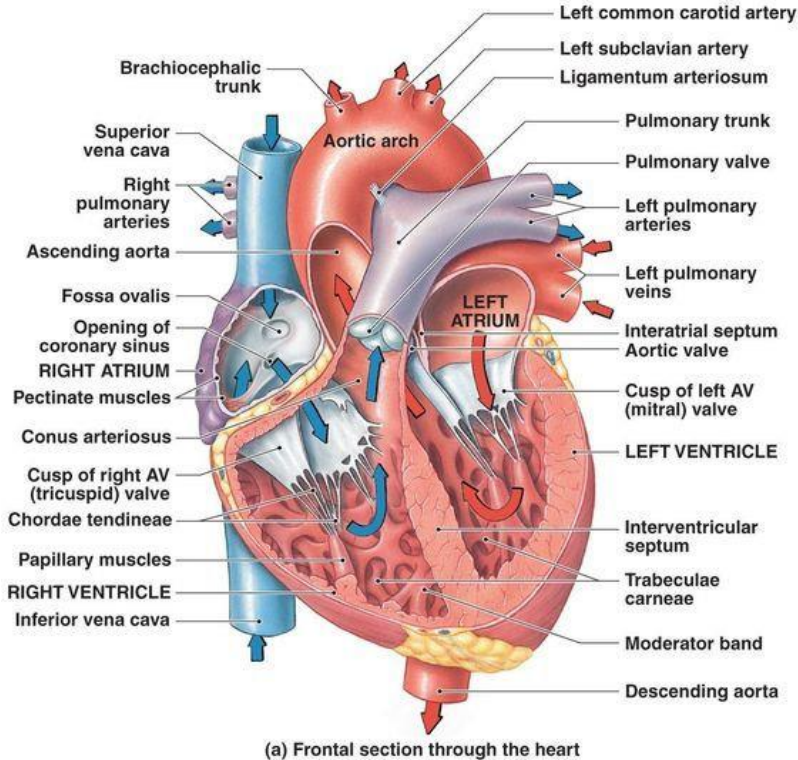


of the estrous cycle phase. Out of the quantified genes, the right side exhibited differential expression patterns. We also revealed also chamber differences in gene expression of ion channels and transporters, and genes with roles in energy metabolism, glycolysis/gluconeogenesis, calcium signaling, and cell cycle regulation. The expression of *Ank2* oscillates in phase with the expression of in-phase oscillating partners in the right ventricle. The analysis indicates that the expression of ion channels and transporters is substantially lower in the right ventricle from the same side.

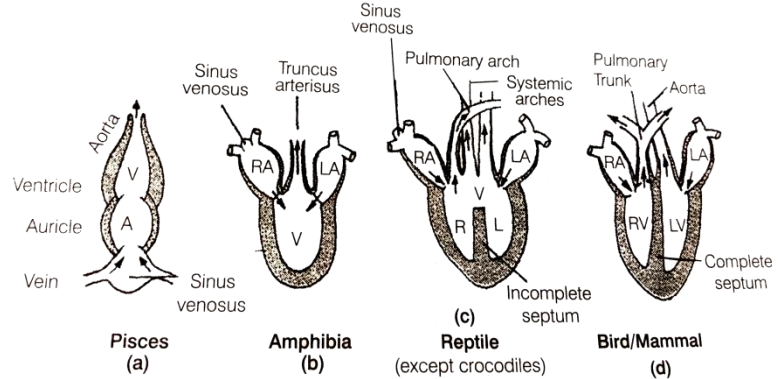


Bucharest

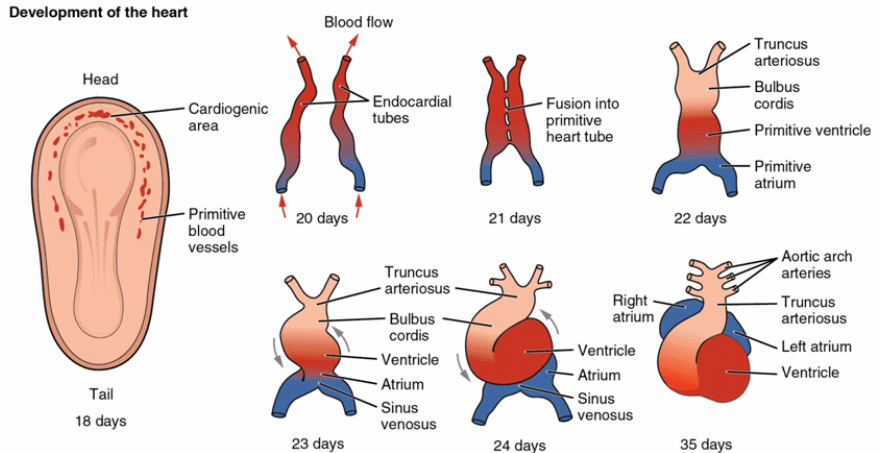
Heart



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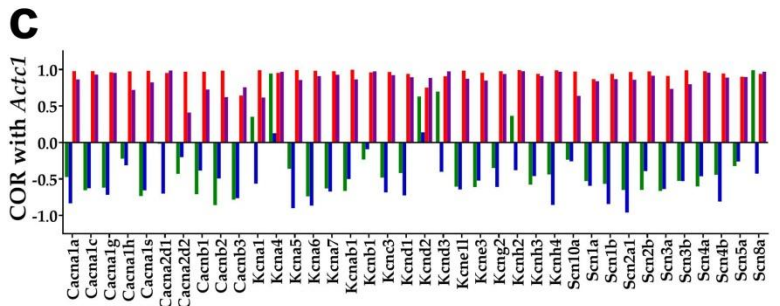
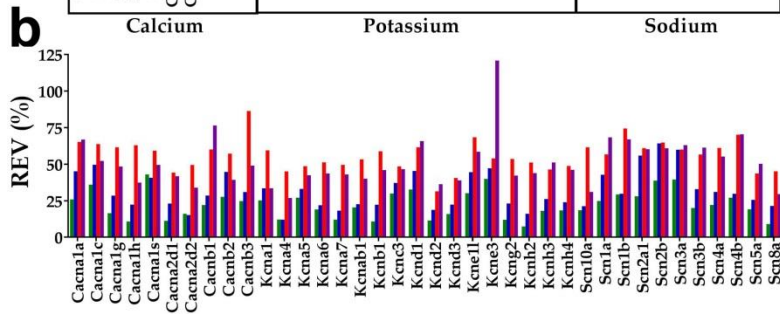
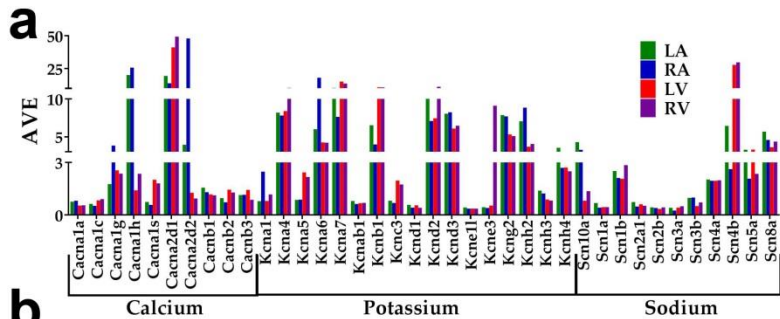


Meckel-Serres Law "Ontogeny recapitulates phylogeny"



★ **Cardiogenic area begins right in the middle of head pole**

Transcriptomic topology



Three independent variables.

Actc1 = actin, cardiac muscle 1

Genomic fabric : $F \equiv (\Gamma, \Pi, \Xi, \Theta)$

Γ = set of composing genes ($|\Gamma|$ = number of genes),

Π = transcriptomic profile (set of the 95% confidence intervals of the composing genes expression levels)

Ξ = control of transcript abundance

Θ = topology (series of many-gene expression correlation functions)

$$AVE_i^{(chamber)} = \frac{1}{R_i} \sum_{k=1}^{R_i} \mu_{i,k}^{(chamber)} = \frac{1}{R_i} \sum_{k=1}^{R_i} \underbrace{\left(\frac{1}{4} \sum_{j=1}^4 a_{i,k,j}^{(chamber)} \right)}_{\mu_i^{(chamber)}}$$

$$REV_i^{(chamber)} = \underbrace{\frac{1}{2} \left(\sqrt{\frac{r_i}{\chi^2(r_i; 1-\alpha/2)}} + \sqrt{\frac{r_i}{\chi^2(r_i; \alpha/2)}} \right)}_{\text{redundancy correction coefficient}} \underbrace{\sqrt{\frac{1}{R_i} \sum_{k=1}^{R_i} \left(\frac{S_{ik}^{(chamber)}}{\mu_{ik}^{(chamber)}} \right)^2}}_{\text{pooled CV}_i^{(chamber)}} \times 100\%$$

$$\Theta(\overline{g_1}, \overline{g_2}, \dots, \overline{g_n}) = \prod_{k=1}^n A_k \left[\overline{F_k(\overline{g_{i1}}, \dots, \overline{g_{ik}})} \mid_{1 \leq i_1 < \dots < i_k \leq n} \right]$$

$$\overline{F_2(\overline{g_{i1}}, \overline{g_{i2}})} \mid_{1 \leq i_1 < i_2 \leq n} = \left(\overline{g_{i1}} r(\overline{g_{i1}}, \overline{g_{i2}}) \overline{g_{i2}} - \overline{g_{i1}} \overline{g_{i2}} \times \hat{1} \right) \mid_{1 \leq i_1 < i_2 \leq n} \quad \text{pair-wise topology}$$

$r(\overline{g_{i1}}, \overline{g_{i2}})$ = Pearson correlation coefficient between the sets of the expression levels of genes i_1 and i_2 within biological replicas

recurrence relation: $\overline{F_k(\overline{g_{i1}}, \dots, \overline{g_{ik}})} \mid_{1 \leq i_1 < \dots < i_k \leq n} = \overline{g_{ik}} r(\overline{g_{ik}}, \overline{F_k(\overline{g_{i1}}, \dots, \overline{g_{ik-1}})} \mid_{1 \leq i_1 < \dots < i_{k-1} \leq n})$, multi dimensional correlations

$$A_1 \geq A_2 \geq A_3 \geq \dots \geq A_n \geq 0 \Rightarrow \Theta(\overline{g_1}, \overline{g_2}, \dots, \overline{g_n}) \approx A_1 \underbrace{\left(\overline{g_{i1}} \times \hat{1} \right)}_{\text{single-gene topology}} \mid_{1 \leq i_1 \leq n} + A_2 \underbrace{\left(\overline{g_{i1}} r(\overline{g_{i1}}, \overline{g_{i2}}) \overline{g_{i2}} - \overline{g_{i1}} \overline{g_{i2}} \times \hat{1} \right)}_{\text{pair-wise topology}} \mid_{1 \leq i_1 < i_2 \leq n}$$

Differential expression

$\forall A, B = RA, LA, RV, LV \quad , \quad A \neq B$

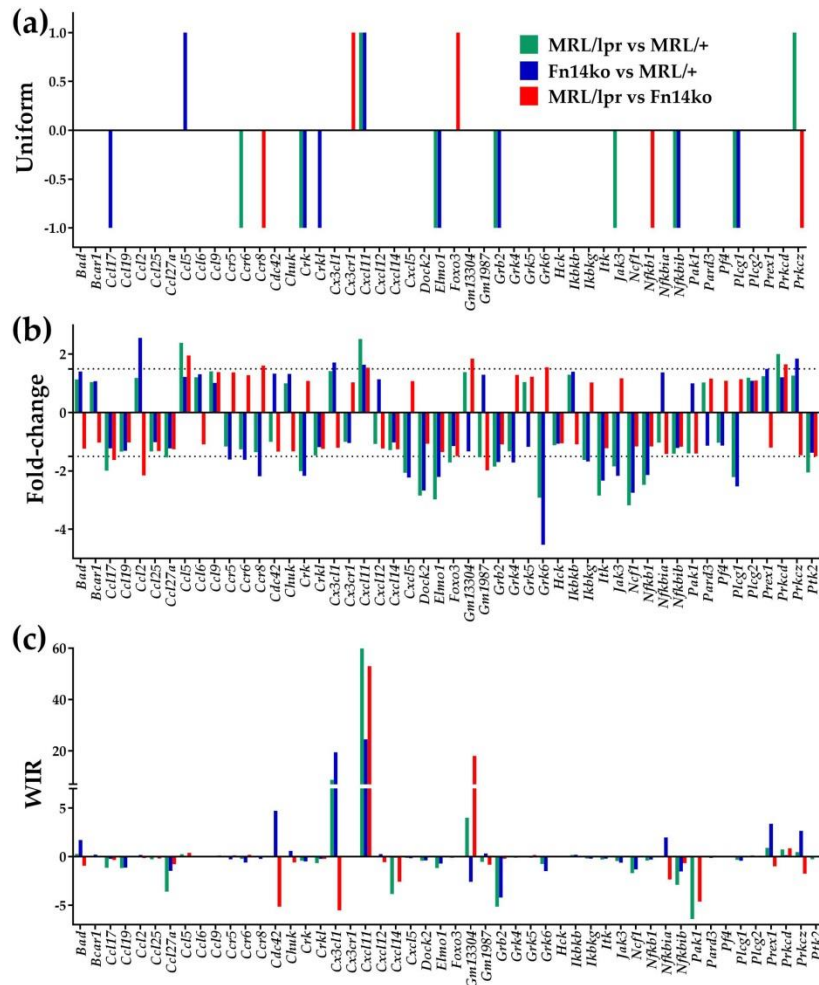
$$x_i^{(A \rightarrow B)} = \begin{cases} \frac{\sum_{k=1}^{R_i} \mu_{ik}^{(B)}}{\sum_{k=1}^{R_i} \mu_{ik}^{(A)}} & \text{if } \sum_{k=1}^{R_i} \mu_{ik}^{(B)} \geq \sum_{k=1}^{R_i} \mu_{ik}^{(A)} \\ -\frac{\sum_{k=1}^{R_i} \mu_{ik}^{(A)}}{\sum_{k=1}^{R_i} \mu_{ik}^{(B)}} & \text{if } \sum_{k=1}^{R_i} \mu_{ik}^{(B)} < \sum_{k=1}^{R_i} \mu_{ik}^{(A)} \end{cases}, \quad \frac{1}{R_i} \sum_{k=1}^{R_i} \mu_{ik}^{(A)} = AVE_i^{(A)}$$

Statistically significant differential expression if:

$$|x_i^{(A \rightarrow B)}| > CUT_i^{(A \rightarrow B)} = 1 + \sqrt{2 \left((REV_i^{(A)})^2 + (REV_i^{(B)})^2 \right)} \quad \wedge \quad p_i^{(A \rightarrow B)} < 0.05$$

Weighted Individual gene Regulation (WIR):

$$WIR_i^{(A \rightarrow B)} \equiv AVE_i^{(A)} \underbrace{\frac{x_i^{(A \rightarrow B)}}{|x_i^{(A \rightarrow B)}|}}_{\text{regulation sign}} \underbrace{\left(|x_i^{(A \rightarrow B)}| - 1 \right)}_{\text{absolute net fold-change}} \underbrace{\left(1 - p_i^{(A \rightarrow B)} \right)}_{\text{confidence of the regulation}}$$



Transcriptomic chamber specificity

$WPR_{\Gamma}^{(A \rightarrow B)} = \overline{wpr_i^{(A \rightarrow B)}}_{i \in \Gamma}$, where:

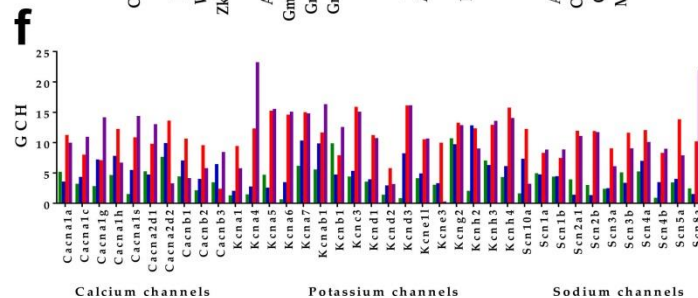
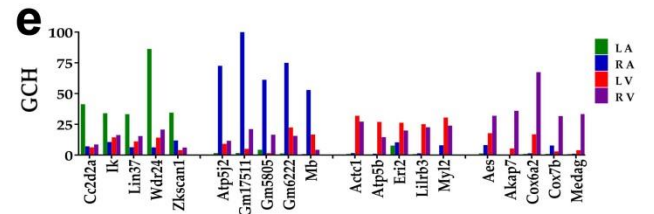
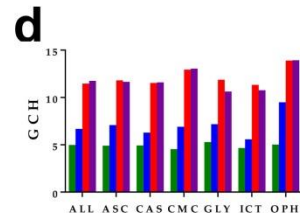
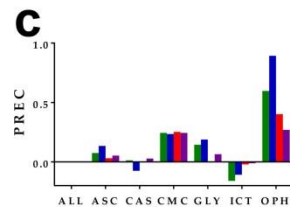
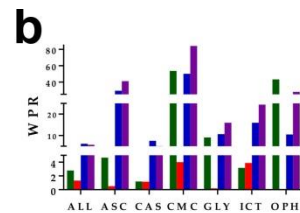
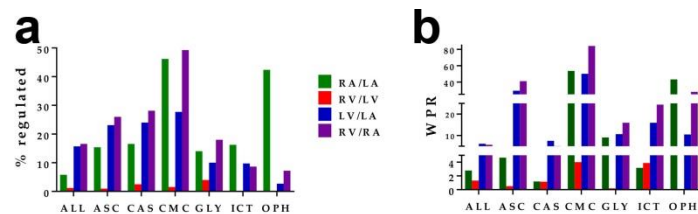
$$wpr_i^{(A \rightarrow B)} = \begin{cases} \mu_i^{(A)} \left(|x_i^{(A \rightarrow B)}| - CUT_i^{(A \rightarrow B)} \right) (1 - p_i^{(A \rightarrow B)}) & \text{if } |x_i^{(A \rightarrow B)}| > CUT_i^{(A \rightarrow B)} \\ 0 & \text{if } |x_i^{(A \rightarrow B)}| \leq CUT_i^{(A \rightarrow B)} \end{cases}$$

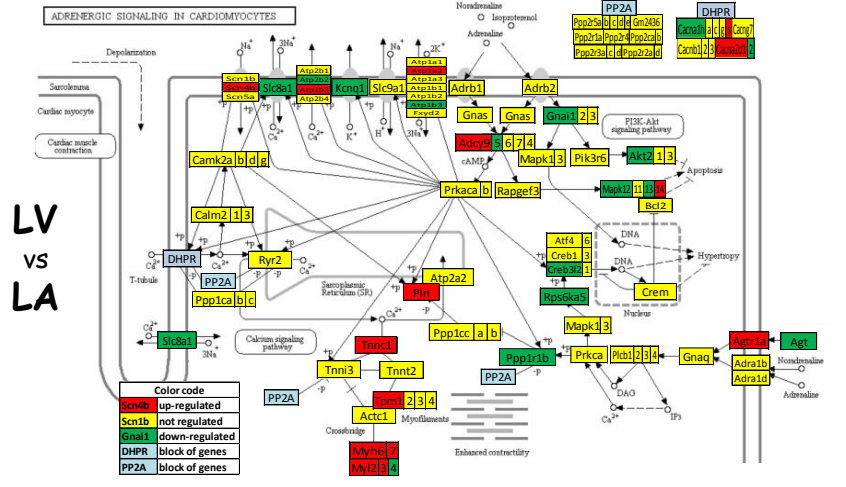
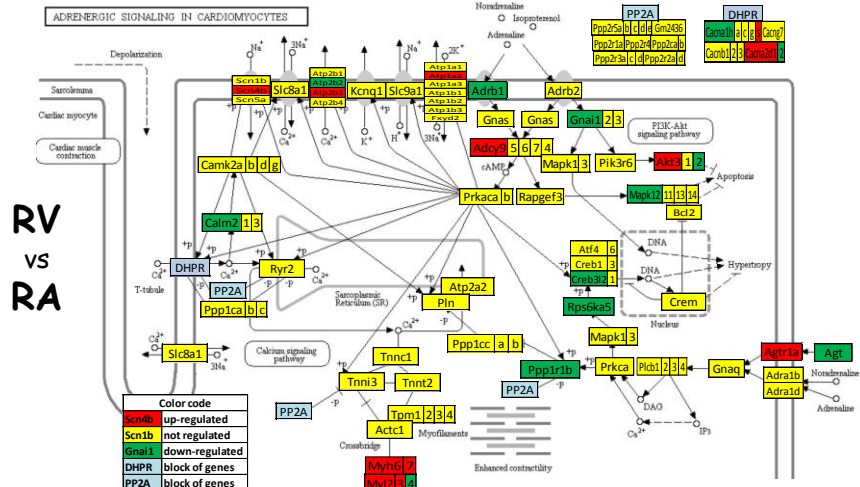
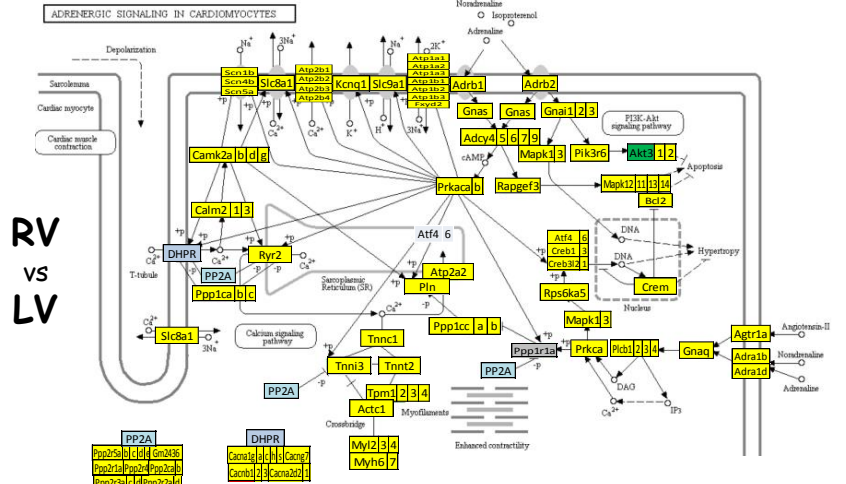
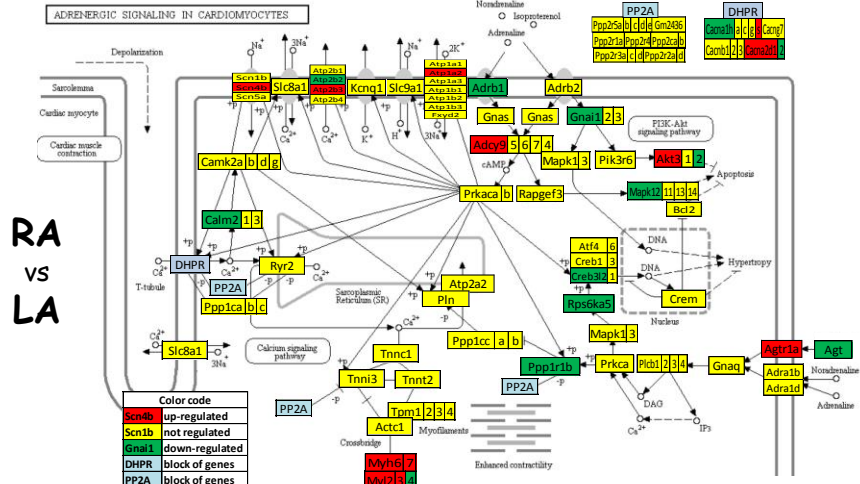
$p_i^{(A \rightarrow B)}$ = p-val of the heteroscedastic t-test of $\mu_i^{(B)} = \mu_i^{(A)}$

$$REC_i^{(chamber)} = \frac{\langle REV^{(chamber)} \rangle_{ALL}}{\langle REV_i^{(chamber)} \rangle} - 1, \quad PREC_{\Gamma}^{(chamber)} = \frac{\langle REV^{(chamber)} \rangle_{ALL}}{\langle REV^{(chamber)} \rangle_{\Gamma}} - 1$$

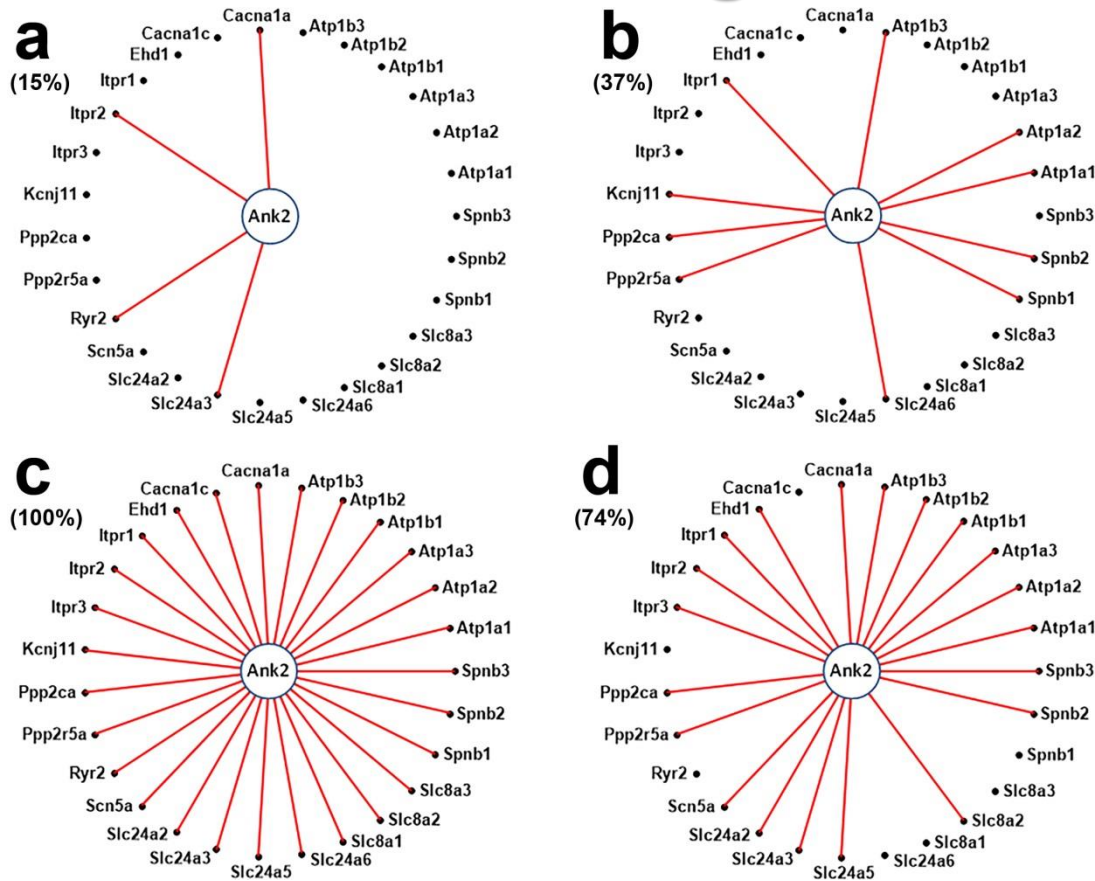
where: $\langle REV^{(chamber)} \rangle_{\Gamma/ALL} \equiv$ median of REV over pathway Γ / entire transcriptome

$$GCH_i^{(chamber)} = (REC_i^{(chamber)} + 1) \times \exp\left(4 \overline{\rho_{ij}^2}^{(chamber)}_{\forall j \neq i}\right)$$



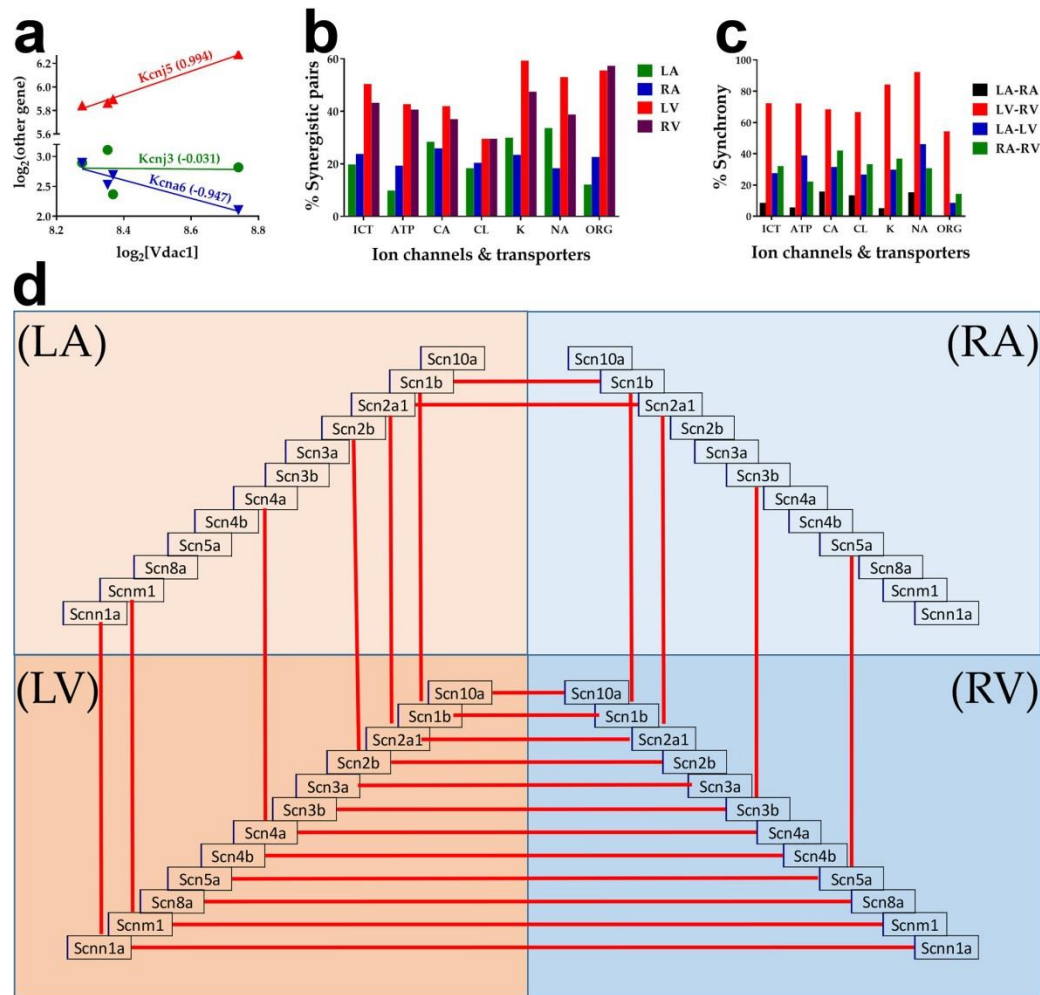


Differential gene networking



Expression coordination of *Ank2* (a major player in cardiac physiology and a hub-bottleneck gene in atrial fibrillation) with its known binding partners in: a. left atrium, b. right atrium, c. left ventricle, d. right ventricle.

Expression synchrony of ionic channels and transporters



Sexual dichotomy

ORIGINAL PAPER

Sex-dependent gene regulatory networks of the heart rhythm

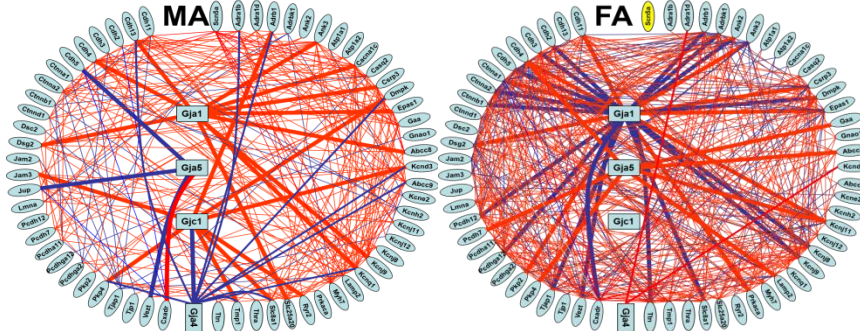
D. A. Iacobs · S. Iacobs · N. Thomas · I. ...



Received: 30 April 2009 / Revised: 19 August 2009 / Accepted online: 16 September 2009
© Springer-Verlag 2009

Abstract Expression level, control, and intercoordination of 66 selected heart rhythm determinant (HRD) genes were compared in atria and ventricles of four male and four female adult mice. We found that genes encoding various adrenergic receptors, ankyrins, ion channels and transporters, connexins, cadherins, plakophilins, and other components of the ...

higher expression in atria than ventricles for males and higher expression in ventricles than atria for females. We have ranked the selected genes according to their prominence (new concept) within the HRD gene web defined as extent of expression coordination with the other web genes and stability of expression. Interestingly, the prominence ...



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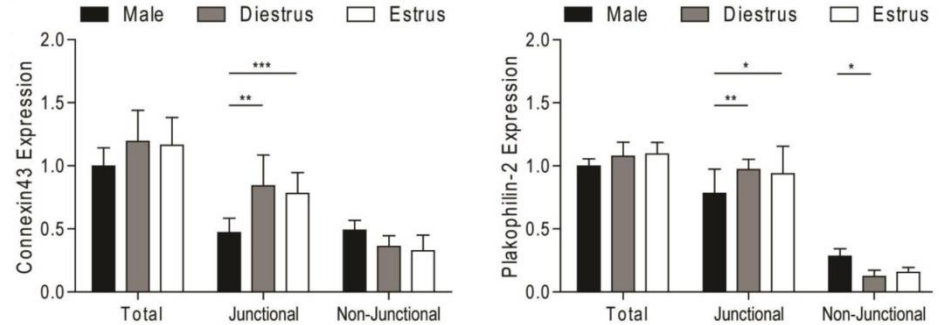
journal homepage: www.elsevier.com/locate/ybbrc



Sex differences in expression and subcellular localization of heart rhythm determinant proteins

N.M. Thomas^{a,*}, J.F. Jasmin^b, M.P. Lisanti^b, D.A. Iacobs^a

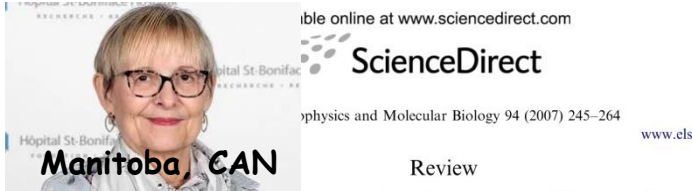
^aDominick P. Purpura Department of Neuroscience, Albert Einstein College of Medicine, 1300 Morris Park Ave., Kennedy Center, New York, NY 10461, USA
^bThomas Jefferson University, Department of Stem Cell Biology and Regenerative Medicine, Bluemle Building, 233 South 10th St., Philadelphia, PA 19107, USA



Gja1 (Cx43), Gja5 (Cx40), Gjc1 (Cx45)

Heart connexins

Physiol Genomics 20: 211–223, 2005.
First published December 7, 2004; doi:10.1152/physiolgenomics.00229.2003.



Available online at www.sciencedirect.com

ScienceDirect

Physiology and Molecular Biology 94 (2007) 245–264

www.elsevier.com

Review

The role of connexins in controlling cell growth and gene expression

Elissavet Kardami^{a,*}, Xitong Dang^a, Dumitru A. Iacobas^b, Barbara E. Nickerson^a,
Madhumathy Jeyaraman^a, Wattamon Srisakuldee^a, Janna Makazan^a,
Stephane Tanguy^a, David C. Spray^b

^aInstitute of Cardiovascular Sciences, University of Manitoba and St. Boniface Research Centre, Winnipeg, Man., Canada

^bDepartment of Neuroscience, Albert Einstein College of Medicine, NY 10461, USA

Available online 16 March 2005



Einstein, NY

Gap Junctions and Chagas Disease

Daniel Adesse,^{*,†} Regina Coeli Goldenberg,[†] Fabio S. Fortes,[‡] Jasmin,^{*,§} Dumitru A. Iacobas,[§]
Sanda Iacobas,[§] Antonio Carlos Campos de Carvalho,^{*,§} Maria de Narareth Meirelles,[†] Huan Huang,[†] Milena B. Soares,^{||} Herbert B. Tanowitz,[†]
Luciana Ribeiro Garzoni,[†] and David C. Spray[§]



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Neurochemistry International 45 (2004) 243–250

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Valhalla, NY

Genes controlling multiple functional pathways are transcriptionally regulated in connexin43 null mouse heart

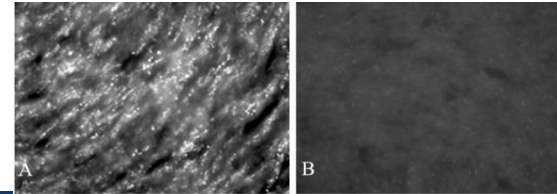
Dumitru A. Iacobas,¹ Sanda Iacobas,¹ W. E. I. Li,¹ Georg Zoidl,³ Rolf Dermietzel,³ and
David C. Spray²

¹Departments of ¹Neuroscience and ²Cardiology, Albert Einstein College of Medicine, Bronx, New York
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Received 15 October 2004; accepted in final form 30 November 2004



Toronto, CAN



Bochum,
GER

Gene expression alterations in connexin null mice extend beyond the gap junction

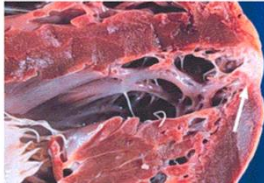
Dumitru A. Iacobas, Eliana Scemes, David C. Spray*

Chagas cardiomyopathy

Parasitol Res (2003) 91: 187–196
DOI 10.1007/s00436-003-0937-z

ORIGINAL PAPER

Hipertrofia



Bronx, NY

Shankar Mukherjee · Thomas J David C. Spray · Dumitru A. Ia Richard N. Kitsis · Murray Wit Philip E. Scherer · Aihao Ding

Microarray analysis of changes in gene expression in a mouse model of chronic chagasic cardiomyopathy

Gene Expression Changes Associated with Myocarditis and Fibrosis in Hearts of Mice with Chronic Chagasic Cardiomyopathy

Milena Botelho Pereira Soares,^{1,2} Ricardo Santana de Lima,¹ Leonardo Lima Rocha,¹ Juliana Fraga Vasconcelos,¹ Silvia Regina Rogatto,^{3,4} Ricardo Ribeiro dos Santos,^{1,2} Sandra Iacobas,⁷ Regina Coeli Goldenberg,⁵ Dumitru Andrei Iacobas,⁷ Herbert Bernard Tanowitz,^{4,9} Antonio Carlos Campos de Carvalho,^{5,6} and David Conover Spray⁸



Chagas I, Brazil

Reversion of gene expression alterations in hearts of mice with chronic chagasic cardiomyopathy after transplantation of bone marrow cells

Milena B.P. Soares,^{1,2,*} Ricardo S. Lima,³ Bruno S.F. Souza,^{1,2} Juliana F. Vasconcelos,^{1,2} Leonardo L. Rocha,¹ Ricardo Ribeiro dos Santos,^{1,2} Sandra Iacobas,⁸ Regina C. Goldenberg,⁹ Michael P. Lisanti,⁹ Dumitru A. Iacobas,¹ Herbert B. Tanowitz,^{6,7,*} David C. Spray^{4,6} and Antonio C. Campos de Carvalho^{4,5,8}



Microbes and Infection xx (2017) 1–11

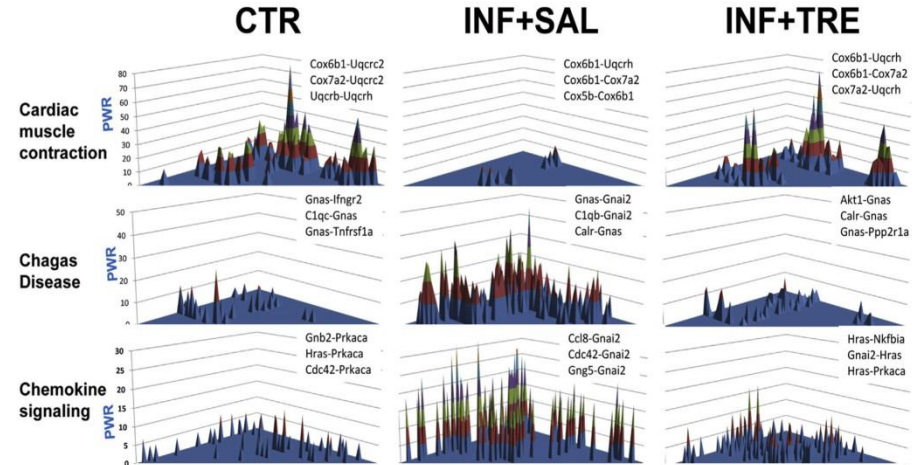


Chagas I, Brazil

Original article

Functional genomic fabrics are remodeled in a mouse model of Chagasic cardiomyopathy and restored following cell therapy

Dumitru A. Iacobas^{a,b,*}, Sandra Iacobas^a, Herbert B. Tanowitz^{c,d}, Antonio Campos de Carvalho^{b,e}, David C. Spray^{b,c}



Original article

Transcriptomic alterations in *Trypanosoma cruzi*-infected cardiac myocytes

Regina Coeli dos Santos Goldenberg^{a,b,c}, Dumitru A. Iacobas^b, Sанда Leonardo Lima Rocha^{a,1}, Fabio da Silva de Azevedo Fortes^{a,b,c}, Leandra Fnu Nagajyothi^a, Antonio Carlos Campos de Carvalho^{b,c}, Herbert B. Tanowitz^a, David C. Spray^{b,*}



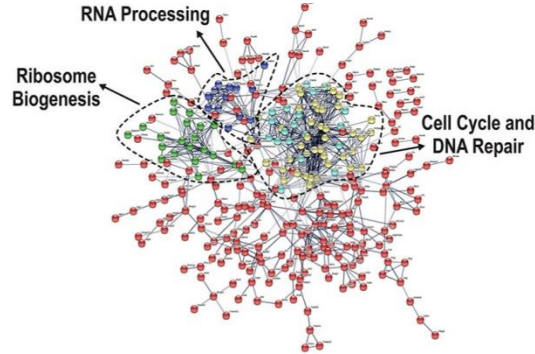
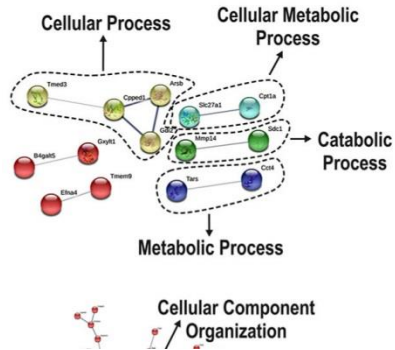
UFRJ, Brazil

Am. J. Trop. Med. Hyg., 82(5), 2010, pp. 846–854
doi:10.4269/ajtmh.2010.09-0399

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Transcriptomic Signatures of Alterations in a Myoblast Cell Line Infected with Four Distinct Strains of *Trypanosoma cruzi*

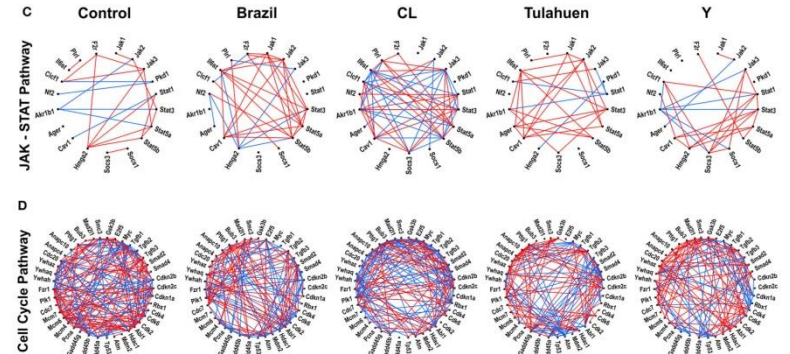
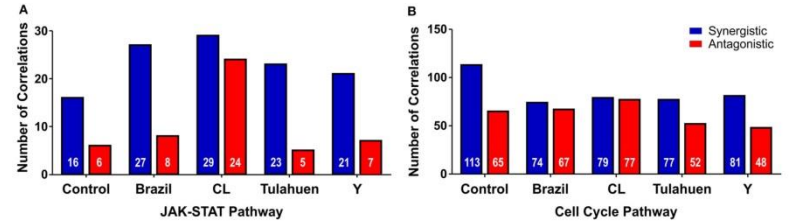
Daniel Adesse,^{*} Dumitru A. Iacobas, Sанда Iacobas, Luciana R. Garzoni, Maria de Nazareth Meirelles, Herbert B. Tanowitz, and David C. Spray



Oswaldo Cruz, BRA

Trypanosoma cruzi Promotes Transcriptomic Remodeling of the JAK/STAT Signaling and Cell Cycle Pathways in Myoblasts

Lindice M. Nishimura¹, Laura L. Coelho², Tatiana G. de Melo³, Paloma de Carvalho Vieira⁴, Pedro H. Victorino⁵, Luciana R. Garzoni⁶, David C. Spray⁷, Dumitru A. Iacobas⁸, Sанда Iacobas⁹, Herbert B. Tanowitz¹⁰ and Daniel Adesse^{11*}



Cardiac ischemia



Fluminense, Brazil



Rio, Brazil

Stem Cell Rev and Rep (2012) 8:251–261

DOI 10.1007/s12015-011-9282-2

Functional and Transcriptomic Recovery of Infarcted Mouse Myocardium Treated with Bone Marrow Mononuclear Cells

Stephan Lachtermacher · Bruno L. B. Esporcatte · Fábio da Silva de Azevedo Fortes · Nazareth Novaes Rocha · Fabrício Montalvão · Patricia C. Costa · Luciano Belem · Arnaldo Rabischoffsky · Hugo C. C. Faria Neto · Rita Vasconcellos · Dumitru A. Iacobas · Sanda Iacobas · David C. Spray · Neil M. Thomas · Regina C. S. Goldenberg · Antonio C. Campos de Carvalho



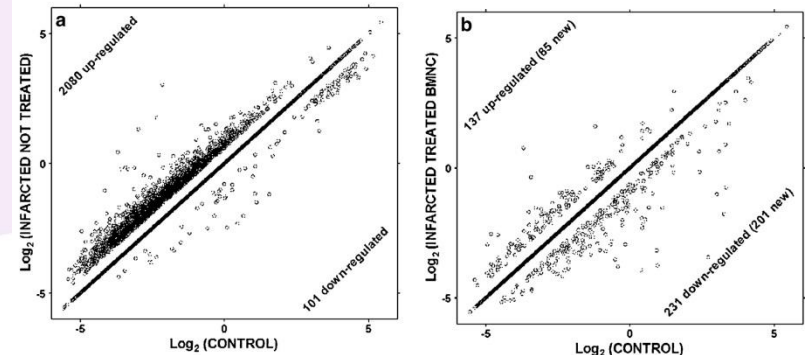
Stem Cell Rev and Rep (2012) 8:251–261

Permanent myocardial infarcts produced by ligation of the descending branch of the left coronary artery BMNC was directly injected using a 10 μ l syringe into 3 different regions (30 μ l total) at the borders of cardiac scar tissue 10 days after experimental infarction.

Braz J Med Biol Res, March 2010, Volume 43(4) 377-389

Cardiac gene expression and systemic cytokine profile are complementary in a murine model of post-ischemic heart failure

S. Lachtermacher, B.L.B. Esporcatte, F. Montalvão, P.C. Costa, D.C. Rodrigues, L. Belem, A. Rabischoffsky, H.C.C. Faria Neto, R. Vasconcellos, S. Iacobas, D.A. Iacobas, H.F.R. Dohmann, D.C. Spray, R.C.S. Goldenberg and A.C. Campos-de-Carvalho



Constant and Intermittent Hypoxia

Physiol Genomics 22: 292–307, 2005.
First published May 31, 2005; 10.1152/physiolgenomics.00217.2004.

Gene expression and phenotypic characterization of mouse heart after chronic constant or intermittent hypoxia

Chenhao Fan,¹ Dumitru A. Iacobas,² Dan Zhou,¹ Qiaofang Chen,¹
James K. Lai,³ Orit Gavrialov,¹ and Gabriel G. Haddad^{1,2}

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Received 15 October 2004; accepted in final form 20 May 2005



Einstein, NY



Pocatello, ID

Integrated transcriptomic response to cardiac chronic hypoxia: translation regulators and response to stress in cell survival

Dumitru A. Iacobas · Chenhao Fan · Sanda Iacobas · Gabriel G. Haddad



San Diego, CA



Biochemical and Biophysical Research Communications

journal homepage: www.elsevier.com/locate/ybbrc

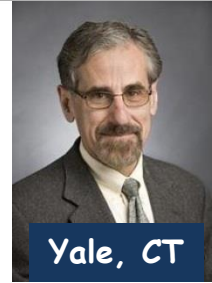
Contents lists available at ScienceDirect



Heart rhythm genomic fabric in hypoxia

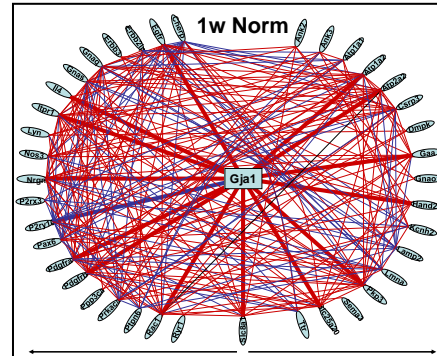
Dumitru A. Iacobas^{a,*}, Sanda Iacobas^a, Gabriel G. Haddad^b

Effects of Chronic Intermittent Hypoxia on Cardiac Rhythm Transcriptomic Networks

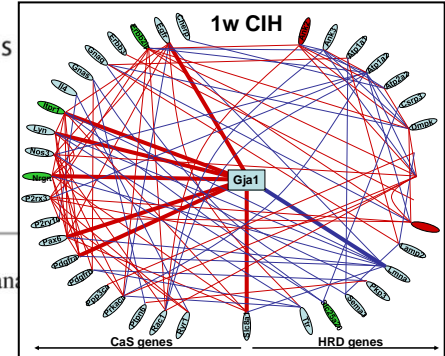


Yale, CT

2



Iacobas



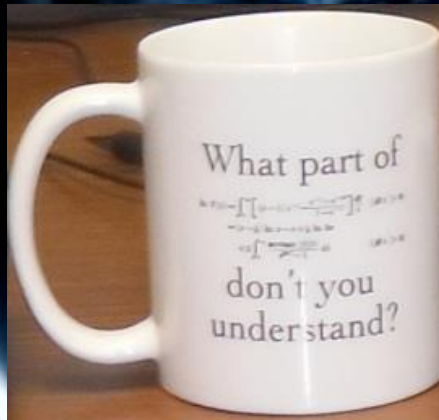
studies and

CaS genes HRD genes

Transcriptomic differences

- LA vs LV > RA vs RV > LA vs RA > LV vs RV in gene expression level, control and coordination
- Male vs female (changes during the estrogen cycle)
- Young vs adult
- Normoxic vs hypoxic (constant or intermittent) or ischemia
- Healthy vs Chagasic cardiomyopathy
- Wildtype vs connexin null/knockdown

Thank you!



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